

## FACT Sheet

## **Recombinant Herpes Viral Vectors**

The following provides information on the use and containment of recombinant herpes viral vectors. Investigators should use these guidelines as part of their risk assessment when planning experiments with these vectors and preparing applications to the Institutional Biosafety Committee (IBC). Note the listed containment levels are the minimum that should be employed with these vectors: some experiments, such as the expression of toxins or oncogenes, may require higher levels of containment. The appropriateness of the containment should be considered as part of the investigator's risk assessment and will be reviewed by the IBC.

NIH Risk Group	RG2
	Herpesviruses are enveloped, icosahedral, double-stranded linear DNA
	viruses.
Biocontainment Level	BSL-2
Infectious to	Yes
Humans/Animals	
Route of Transmission	HSV-1 is typically transmitted by saliva or by the infection on hands of
	healthcare personnel. HSV-2 is typically transmitted through sexual contact.
	HSV can be transmitted by direct contact with epithelial or mucosal surfaces.
Laboratory Hazards	In the laboratory, HSV can be transmitted by ingestion, parenteral injection,
	droplet exposure of the mucous membranes (eyes, nose or mouth), and
	inhalation of aerosolized materials.
Disease	Depends on type:
	Oral Herpes
	Genital Warts
	Herpes esophagitis
	Herpes encephalitis or meningitis
Treatment/Prophylaxis	Antivirals may reduce shedding
Pathogenesis	After infection, the viruses are transported along sensory nerves to the nerve
	cell bodies, where they reside lifelong. Causes of recurrence may include:
	decreased immune function, stress, and sunlight exposure. The first episode
	is often more severe and may be associated with fever, muscle pains, swollen
	lymph nodes and headaches. Over time, episodes of active disease decrease
	in frequency and severity

<b>Replication Competent</b>	All versions of HSV vectors are prone to recombination. Additionally,
	approximately 50% - 90% of adults possess antibodies to HSV type 1; 20% -
	30% of adults possess antibodies to HSV type 2. This is a concern since
	reactivation from latency is not well understood. Infection by HSV vectors
	into latently infected cells could potentially reactivate the wild-type virus, or
	spontaneous reactivation of a latent infection could produce an environment
	where replication defective vectors could replicate.
RCV Testing	Viral preparations used for in vitro studies should be tested every 6 months
	for replication competent viruses by plaque assay. These assays should be
	tested at a sensitivity limit of 1 infectious unit per mL.
Disinfection	Effective disinfectants require a minimum of 20 minutes contact time. Use
	one of the following:
	RECOMMENDED: Sodium hypochlorite (0.5%: use 1:10 dilution of fresh
	bleach)
	• 5% Phenol
	70% Ethanol or Isopropanol
Animals	ABSL-2: Animals will be maintained at ABSL-2 for the duration of the study.
	Animals must be injected in a Biological Safety Cabinet. All bedding, waste
	and animals infected with HSV shall be treated as biohazardous. After all
	animals are removed from their primary enclosure immediately autoclave or
	treat with chemical disinfectant. After disinfection, dump the cage contents
	and begin cleaning the cage for re-use. All waste must be decontaminated by
	autoclaving or chemical disinfection prior to disposal. Animal carcasses must
	be placed in autoclave bags and be designated for infectious waste disposal.
	All necropsies must be performed in a designated room using animal BSL-2
	practices and procedures.
	Animal cages must be labeled with a biohazard sign.

## Sources:

http://web.stanford.edu/dept/EHS/prod/researchlab/bio/docs/Working with Viral Vectors.pdf http://www.dartmouth.edu/~ehs/biological/biosafety\_docs/110\_1\_ibc\_viral\_vector\_policy.pdf